

Preparation and characterization of potato starch/bentonite organophylized by *SDS*

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This study focuses on the development of nanocomposite films using potato starch via a casting technique. Glycerol was incorporated as a plasticizer, while Sodium Dodecyl Sulfate (*SDS*)-modified bentonite acted as reinforcement. Various analytical methods, including Fourier transform infrared (FTIR), X-ray diffraction (XRD), Thermogravimetric analysis (TGA), UV/vis spectroscopy, Electron microscopy (SEM), and Optical microscopy (MOP), were utilized to assess the material properties. Different concentrations of *SDS*, below, at, and above the critical micelle concentration (*CMC*), were investigated. The findings suggest that films containing higher *SDS* levels demonstrate enhancements in both physical and chemical characteristics. XRD assessments reveal that films with *SDS* concentrations surpassing the *CMC* level display homogeneity, indicating effective intercalation or exfoliation processes. This compatibility of the biofilm components is further evidenced through FTIR, SEM, and MOP. Moreover, the introduction of *SDS* significantly improves the film's biodegradability, as confirmed by thermogravimetric analysis.

Keywords: Biofilms; starch; bentonite; *SDS* surfactant; compatibility.

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1. Introduction

While petroleum-based plastic products have become integral to various aspects of daily life, their poor degradability poses a significant environmental threat [1–3]. Consequently, considerable research efforts are being directed towards tackling the challenges of plastic waste and advocating for the large-scale adoption of eco-friendly alternatives. Polysaccharides, like starch derived from potatoes or rice, emerge as promising substitutes for traditional plastics due to their abundance, renewability, and cost-effectiveness [4, 5]. Although polysaccharide-based plastics hold promise, they face inherent physical and chemical limitations, prompting researchers to explore composite materials to improve their properties [6–8]. Indeed, some studies have shown that the addition of carefully chosen components significantly improves the physicochemical and mechanical properties of these biofilms. For example, the incorporation of glycerol, functioning as a plasticizer, bestows flexibility upon the material by intercalating amidst starch molecules, resulting in more supple films [9–11]. Additionally, the integration of bentonite clay notably bolsters mechanical strength owing to its distinctive chemical composition and structure [12, 13]. Bentonite clay typically comprises multiple layers with interlayer spaces hosting regularly packed ions. Under conducive conditions, the interaction between bentonite and charged molecules prompts intercalation and, in certain instances, exfoliation, thereby augmenting the uniformity and mechanical prowess of the composite [14, 15].

On the other hand, the organophilisation of bentonite, achieved through modification with organic molecules like surfactants, seeks to impart hydrophobic properties to the bentonite, thereby easing its integration into organic polymers. In addition to enhancing dispersion and mechanical reinforcement, this versatile technique enables efficient adsorption and removal of contaminants. Moreover, the expanded application range resulting from organophilisation greatly enhances the performance of composite materials, benefiting various industrial sectors.

This investigation zeroes in on enhancing starch/glycerol-based films reinforced with bentonite and Sodium Dodecyl Sulfate (*SDS*) surfactant, renowned for its beneficial attributes. Specifically, we delve into the impact of *SDS* concentration, scrutinizing three concentrations—higher, equivalent, and lower than its critical micellar concentration—analyzing the ensuing outcomes.

2. Materials and methods

2.1. Materials

The chemicals used in our experiments are commercially available products. The potato-starch (Stch), a white fine powder with 10 wt% moisture content containing 35 – 15 wt% amylose and 65 – 85 wt% amylopectin, was purchased from Tuoketuohua Starch Company (Neimeng, China). The Bentonite (Bt) and glycerol were obtained from Sigma-Aldrich Co. The structural formula of the anionic surfactant, sodium dodecyl sulphate (*SDS*) is

($C_{12}H_{25}SO_4^- + Na^+$), was purchased from FlukaChemical. All chemicals used are pure grade.

2.2. Experimental techniques and characterization

2.2.1. X-ray diffraction analysis

X-ray diffraction was conducted using a Mini Flex 600 X-ray diffractometer to measure inter-reticular distance and determine the exfoliation rate. The following conditions were employed: F.F tube 40 kV and 15 mA. Films were exposed to Cu radiation at a wavelength of 1.54 nm and scanned over a 2θ diffraction angle interval of $2 - 30^\circ$ with a scan speed of $10^\circ/\text{min}$.

2.2.2. Transform infrared (FTIR) spectra

FTIR spectra of the films were recorded using an Agilent Cary 640 FTIR spectrophotometer across the range of $400\text{--}4000\text{ cm}^{-1}$ employing a transmission method.

2.2.3. Thermal properties

TGA-DTA analysis of films was performed using a Linseis TGA PT1600 differential thermal analyzer. Samples weighing 9-10 mg were heated to 600°C at a rate of $10^\circ/\text{min}$ under a nitrogen atmosphere. Weight loss was measured as a function of temperature.

2.2.4. Opacity

Opacity values of the films were determined using an Analytikjena, Specord 200 plus UV/VIS spectrophotometer. Rectangular pieces of films were placed in the sample compartment of the spectrophotometer. An empty compartment served as a reference in the measurements. Absorbance spectra of the films were recorded over the range of 200 to 800 nm, and opacity values were calculated using the equation:

$$\text{Opacity} = \text{Abs}(600)/X, \quad (1)$$

where Abs 600 is the absorbance value at 600 nm and X (mm) is the film thickness.

2.2.5. Scanning electron microscope

The surface morphology of *Stch/Bt* and *Stch/SDS/Bt* films was examined using a scanning electron microscope performed with a VEGA3 TESCAN. Computations were made using Microcal Origin.

2.2.6. Optical microscope

Optical microscopy (MOP) analysis was conducted using an OLYMPUS BX41.

2.2.7. Measurement of film thickness

Film thickness was measured using an electronic outside micrometer with a sensitivity of 0.001 mm. The average of ten measurements for each thickness was considered in this work.

2.3. Film preparation

Four solutions of bentonite with a concentration of 0.125 g per 45 g of water were prepared using magnetic stirring for a duration of 24 hours, followed by sonication for one hour. To achieve three different concentrations, *SDS* was incorporated into the previously prepared bentonite solutions, resulting in higher (0.0035 g/ml), lower (0.0018 g/ml), and equal (0.0025 g/ml) concentrations compared to the CMC. These three solutions were further stirred for an additional 24 hours. Additionally, potato starch (2.5 g) and glycerol (0.75 g) were added to each of the three previously prepared solutions, which were subsequently subjected to mechanical agitation for 15 minutes at a temperature of 80°C . A fourth control solution was prepared without the addition of *SDS* using the same methodology. All solutions, prepared at a *pH* of 5.5, were poured into Petri dishes and dried for a week at room temperature. The resulting dry films were then removed from the molds.

3. Results and discussions

3.1. Representation of the different compositions of the solutions

The images presented in Fig. 1a) illustrate that the film, in the absence of *SDS*, exhibits fragility and lacks visual appeal. Films produced at *SDS* concentrations equal to or below its Critical Micelle Concentration (*CMC*) are prone to deterioration under storage conditions, as indicated by the presence of spots on the film surface. Conversely, when *SDS* concentration surpasses the *CMC* threshold [as illustrated in Fig. 1c)], the film achieves its optimal condition.

TABLE I. Representation of the different compositions of the solutions prepared for the films produced.

FILMS COMPOSITION	<i>Bt</i> (g)	<i>SDS</i> (g)	<i>Starch</i> (g)
<i>Bt/Stch</i>	0.125	-	2.5
<i>Bt/SDS</i> _{<CMC}	0.125	0.085	-
<i>Bt/SDS</i> _{=CMC}	0.125	0.1165	-
<i>Bt/SDS</i> _{>CMC}	0.125	0.1581	-
<i>Bt/SDS</i> _{<CMC} / <i>Stch</i>	0.125	0.085	2.5
<i>Bt/SDS</i> _{=CMC} / <i>Stch</i>	0.125	0.1165	2.5
<i>Bt/SDS</i> _{>CMC} / <i>Stch</i>	0.125	0.1581	2.5

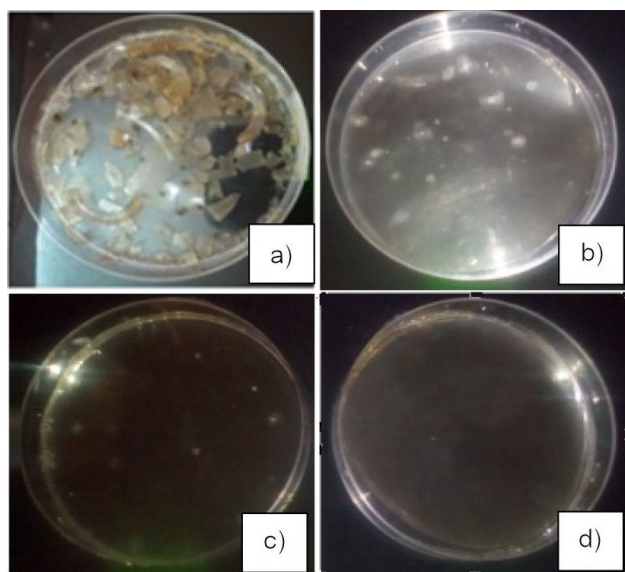


FIGURE 1. Photographs of the different films obtained a) *Bt/Stch*, b) *Bt/SDS*_{<CMC}/*Stch*, c) *Bt/SDS*_{=CMC}/*Stch* and d) *Bt/SDS*_{>CMC}/*Stch*.

3.2. FTIR analysis

Figure 2 showcases the FTIR spectra of *Bt*, *SDS*, and the organo-bentonite *Bt/SDS* composite.

In the *Bt* spectrum, a prominent peak emerges from Al-OH at 915 cm⁻¹, accompanied by a robust peak at 1001 cm⁻¹ attributed to the Si-O bond [16]. Notably, peaks at 3623 and 1633 cm⁻¹ signify stretching vibrations of OH groups in H₂O and the deformation band of OH in the water retained within the silica matrix, respectively [17, 18].

In the standard *SDS* spectrum, stretching vibration peaks of CH₃ and CH₂ groups manifest at 2955 cm⁻¹, 2916 cm⁻¹, and 2849 cm⁻¹, alongside the characteristic absorption peak of the sulfate acid group at 1216 cm⁻¹. Additionally, a pronounced peak at 1467 cm⁻¹ denotes the deformation band of C-H [19, 20].

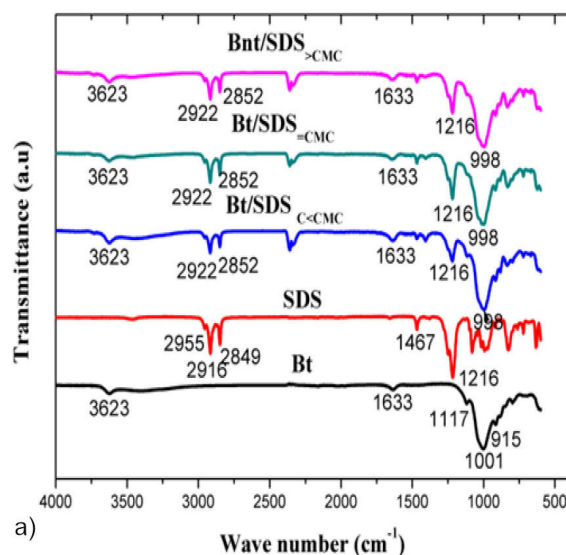
The organo-bentonite spectra of *Bt/SDS* exhibit a slight shift in the vibration peak of the Si-O bond to 998 cm⁻¹, along with characteristic peaks of *SDS* at 1216 cm⁻¹, 2922 cm⁻¹, and 2852 cm⁻¹, suggesting an interaction between *SDS* and bentonite [21].

Figure 2b) depicts the distinct bands of the various film constituents without alterations in peak occurrences, indicating a lack of bands among the diverse *Bt/SDS/Stch* constituents, possibly due to the limited amount of *Bt* employed.

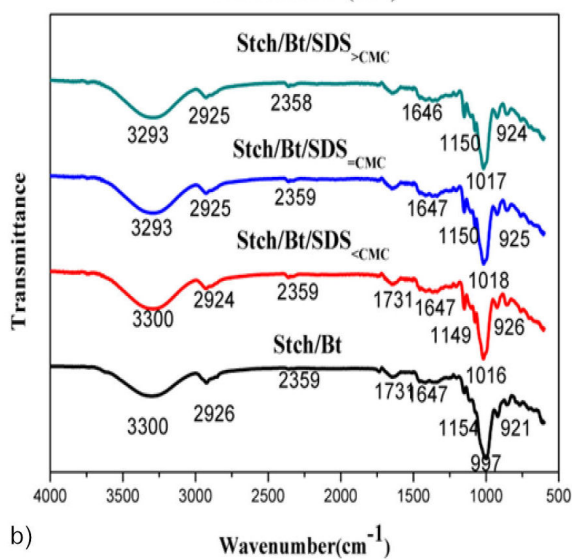
3.3. DRX analysis

The X-ray diffraction patterns of *Bt*, *SDS*, and their three composites are illustrated in Fig. 3.

Bt displays its characteristic peak at $2\theta = 6.8^\circ$, corresponding to the d_{001} basal reflection of the clay. This peak signifies a basal spacing of 12.99 Å, indicating intercalation with a monolayer of water. *SDS* XRD spectra exhibit



a)



b)

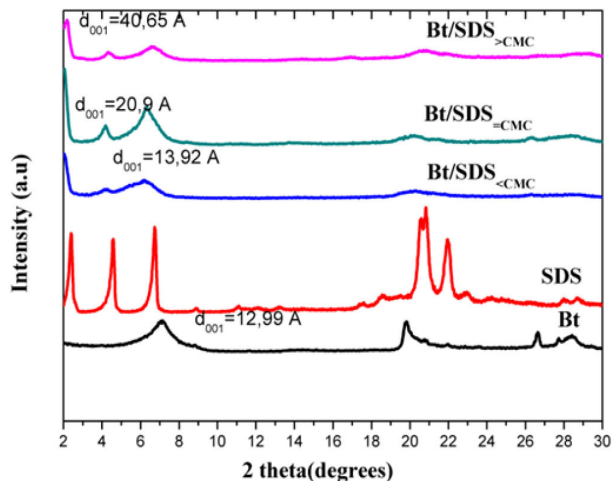
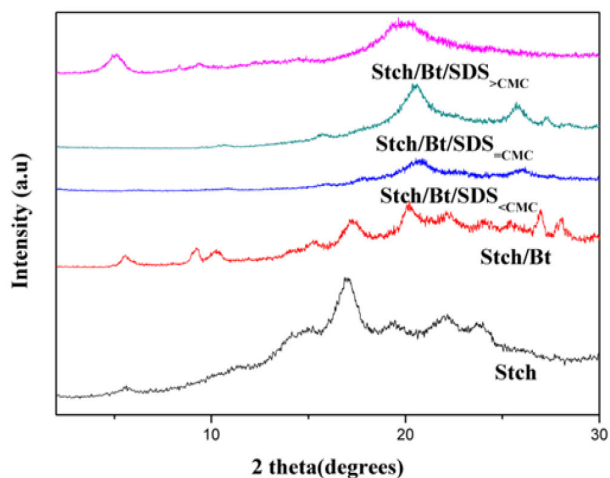
FIGURE 2. FTIR spectra of the *Stch*-based biofilm with the three *SDS* concentrations. a) *Stch/SDS* b) *Stch/Bt/SDS*.

diffraction peaks at $2\theta = 2.406^\circ$, 4.583° , and 6.739° . In the XRD pattern of the *Bt/SDS* composite, the typical *Bt* reflection ($2\theta = 6.8^\circ$) vanishes, replaced by new group reflections. The first distance spacing ($2\theta = 6.35^\circ, 4.23^\circ, 2.172^\circ$) of the composite with varying *SDS* concentrations results in values of 13.92 Å, 20.9 Å, and 40.65 Å, respectively, with a shift towards smaller angles for *Bt/SDS* at *CMC*. FTIR analysis suggests that *SDS* permeates the *Bt* layers, thereby expanding the interlayer space [22, 23]. These observations align with previous literature reports [24].

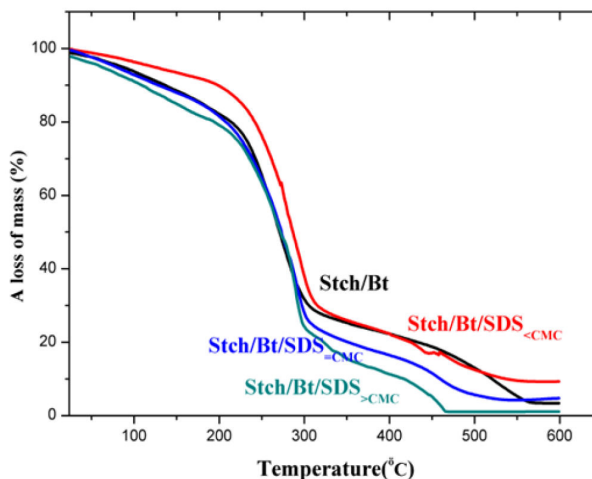
The distinctive peaks of the *Stch/Bt/SDS* films at lower concentrations ($C < CMC$ and $C = CMC$) nearly vanish in Fig. 4, suggesting the presence of exfoliated nanostructures. This phenomenon aligns with previous studies [25, 26]. Conversely, concentrations exceeding the *CMC* reveal pronounced intercalation. Ideally, achieving perfectly

TABLE II. TGA of *Stch/Bt* and different *Stch/Bt/SDS* films-residual mass at different temperatures.

Temperature (°C)	% Δm <i>Stch/Bt</i>	% Δm <i>Stch/Bt/SDS</i> _{<CMC}	% Δm <i>Stch/Bt/SDS</i> _{=CMC}	% Δm <i>Stch/Bt/SDS</i> _{>CMC}
25-100	5.06	3.54	6.76	6.84
100-200	11.62	6.51	11.25	11.91
200-300	50.40	51.78	53.53	54.85
300-400	9.48	15.72	11.55	12.95
400-500	9.24	9.80	10.79	-
500-600	9.66	3.23	0.95	-

FIGURE 3. XRD patterns of *Bt*, *SDS*, *Bt/SDS*_{<CMC}, *Bt/SDS*_{=CMC}, *Bt/SDS*_{>CMC}.FIGURE 4. DRX patterns of *Stch*, *Stch/Bt*, *Stch/Bt/SDS*_{<CMC}, *Stch/Bt/SDS*_{=CMC}, *Stch/Bt/SDS*_{>CMC}.

intercalated or exfoliated nanostructures is necessary for enhancing the performance of polymer nanocomposites [27, 28].

FIGURE 5. TGA curves of *Stch/Bt*, *Stch/Bt/SDS*_{<CMC}, *Stch/Bt/SDS*_{=CMC}, *Stch/Bt/SDS*_{>CMC}

3.4. TGA analysis

Thermogravimetric analysis (TGA) is an important analytical technique for studying biofilms. It allows the determination of the thermal stability and composition of biofilms by measuring the variation in their mass as a function of temperature. This information is essential for understanding thermal degradation, decomposition processes, and the water and organic and inorganic content of biofilms.

In Fig. 5, all intervals have to be considered. Between 25°C and 200°C, for all three curves, there is a monotonous decrease in weight loss, suggesting the progressive release of water or volatile compounds present in the biofilm. The slight inflection point around 100°C could correspond to minor thermal transitions in the biofilm components, such as the desorption of adsorbed water or other volatile compounds [29].

Around 200°C, a sharp drop is observed in all three curves. This could indicate the beginning of thermal decomposition of the biofilm components, such as plasticized starch with glycerol, bentonite, and *SDS*. This abrupt drop suggests a significant transition in the thermal reactions of the biofilm [30, 31].

Between 200°C and 300°C, weight loss continues significantly in this temperature range. The higher the concen-

tration of *SDS*, the greater the weight loss appears to be. This could be due to a more significant decomposition of the biofilm components at higher concentrations of *SDS* [32].

Between 300°C and 450°C, the curve continues to decline, indicating ongoing decomposition of the remaining biofilm components. The observed difference in weight loss between different concentrations of *SDS* could reflect variations in the composition and thermal stability of the biofilm.

Above 450°C, the curves seem to reach a plateau, suggesting that the majority of the biofilm compounds have decomposed or volatilized. This plateau may indicate the presence of stables residues or decomposition products that remain in the system until higher temperatures [33].

These results show the influence of the concentration of *SDS* on the thermal decomposition of the biofilm, with higher concentrations of *SDS* leading to greater weight loss in the studied temperature range. This is due, to the *SDS* whose molecules are easily degradable.

3.5. Opacity and UV analysis

In essence, the intricate interplay between the internal and surface microstructure of a film plays a pivotal role in defining its optical properties, rendering the analysis of opacity a compelling metric for evaluating its suitability in packaging applications. Beyond mere optical considerations, such analysis serves as a window into the underlying structural nu-

ances of film compositions. Moreover, the acknowledgment of thickness variations as a critical determinant of mechanical and barrier characteristics underscores the imperative of meticulous control over processing parameters to uphold quality standards. This holistic understanding, encapsulated in the comprehensive presentation of results in Table III, underscores the multifaceted nature of film assessment and its implications for diverse industrial applications.

Figure 6 presents the UV-visible absorbance spectra of two types of matrices: *Stch/Bt* and *Stch/Bt/SDS*. The films used for these measurements were adjusted to a thickness ranging between 0.5 and 0.75 mm.

In the graph, we observe a slight alteration in the optical clarity of the Starch/Bentonite matrix film when *SDS* is introduced. With an increase in *SDS* concentration within the Starch/Bentonite matrix, there's a corresponding rise in absorbance across the visible UV spectrum [34]. Notably, the Starch/Bentonite/*SDS* matrix exhibits the highest absorbance within the 200-400 nm range compared to all other samples. This heightened absorbance can be attributed to the effective and uniform dispersion of the *SDS* surfactant within the Starch/Bentonite matrix [35].

3.6. SEM analysis

The dispersion and distribution of surfactants in composites, assessed via scanning electron microscopy (SEM), are vital for determining composite properties. SEM provides detailed images that show how uniformly the reinforcement material is spread and distributed within the polymer matrix.

Figure 7 presents images showcasing the morphological disparities between composites with and without *SDS*. Specifically, in images b) and c) spotlight the profound impact of compatibility and homogeneity on the properties of the films under investigation. It is noteworthy that any white spots observed in these images, such as spherical or lenticular shapes, represent starch granules stemming from the precipitation and fermentation process of the starch [36]. These observations provide valuable insights into the structural integrity and composition of the composites, elucidating key factors influencing their performance.

3.7. MOP analysis

To comprehensively analyze the morphology of the produced films, we employed optical measurements microscopy (MOP) to investigate how the addition of *SDS* affects the surface characteristics across varying *SDS* concentrations, as depicted in Fig. 8.

In Fig. 8, image a), clusters and aggregated particles are evident, likely stemming from the poor compatibility of the Starch/Bentonite mixture. Conversely, images b and c vividly exhibit improved compatibility, indicative of the exfoliation effect, particularly noticeable at *SDS* concentrations less than or equal to the critical micelle concentration (CMC) [37]. Figure 8d) presents a distinct appearance compared to

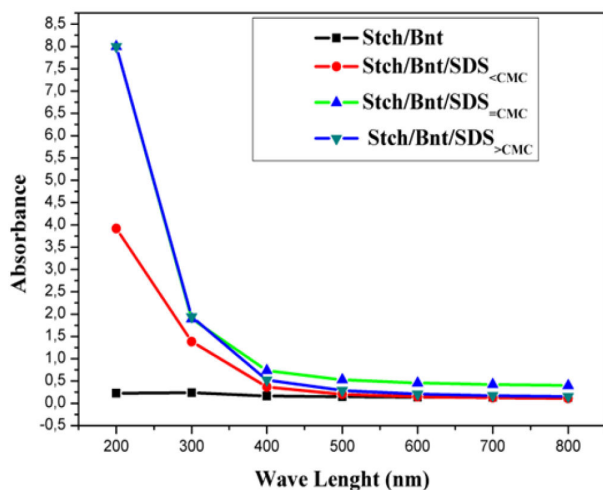


FIGURE 6. Films opacity of *Stch/Bt*, *Stch/Bt/SDS*_{_{C_{MC}}, *Stch/Bt/SDS*_{=C_{MC}}, *Stch/Bt/SDS*_{>C_{MC}}.}</sub></sub>

TABLE III. Measurements carried out for the four films studied.

The sample	Opacity	Thickness (mm)	Opacity/thickness (mm ⁻¹)
<i>Bt/Stch</i>	0.517	0.744	0.694
<i>Bt/SDS</i> _{<sub>C<sub>MC</sub></sub>/<i>Stch</i>}	0.343	0.426	0.805
<i>Bt/SDS</i> _{=C<sub>MC</sub></sub>/<i>Stch</i>}	0.327	0.501	0.652
<i>Bt/SDS</i> _{>C<sub>MC</sub></sub>/<i>Stch</i>}	0.213	0.544	0.393

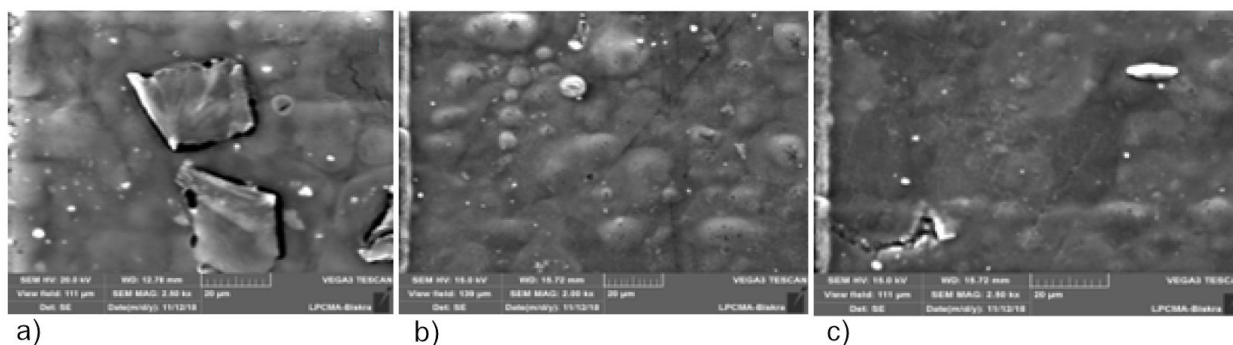


FIGURE 7. SEM images of based starch films with (a) *Stch/Bt*, (b) *Stch/Bt/SDS=CMC*, (c) *Stch/Bt/SDS>CMC*.

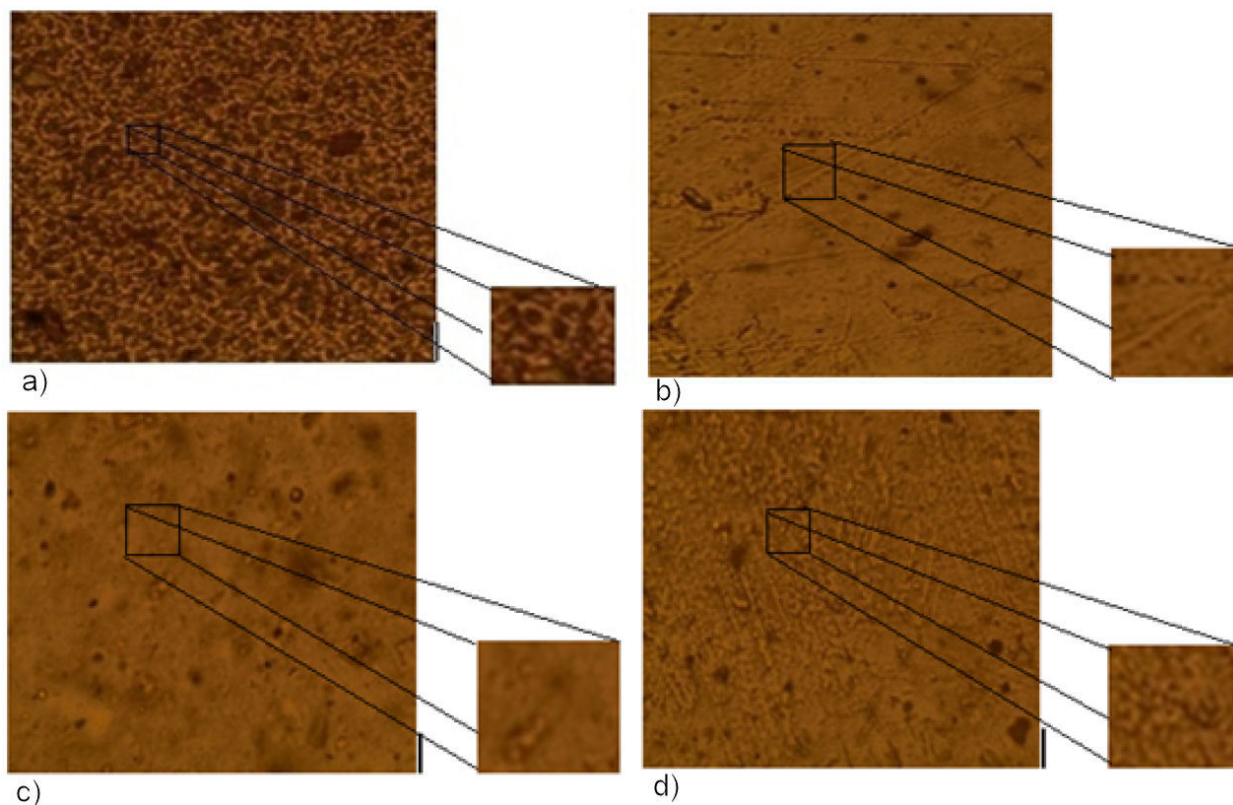


FIGURE 8. MOP images of based starch films with a) *Stch/Bt*, b) *Stch/Bt/SDS<CMC*, c) *Stch/Bt/SDS=CMC*, d) *Stch/Bt/SDS>CMC*.

images a), b), and c), suggesting an intermediate level of compatibility, wherein strong intercalation of *SDS* within the Bentonite matrix is observed. These observations shed light on the nuanced interplay between *SDS* concentration and film morphology, providing valuable insights into the structural evolution of the composites [38].

4. Conclusion

In summary, this investigation underscores the pivotal role of sodium dodecyl sulfate (*SDS*) concentration in shaping the production and properties of biodegradable biofilms. Through an array of meticulous physico-chemical analyses,

it elucidates the intricate interplay between *SDS* and the constituent elements of the biofilm, particularly starch and glycerol-plasticized bentonite. The findings reveal a nuanced relationship wherein low *SDS* concentrations facilitate exfoliation, while higher concentrations induce strong intercalation, resulting in notable enhancements in biofilm homogeneity. Microscopic observations and mass loss analyses further corroborate these insights, shedding light on the mechanisms underlying *SDS*-mediated degradation. Ultimately, this study not only underscores the significance of *SDS* concentration in biofilm production but also heralds promising avenues for the advancement of sustainable and eco-friendly materials.

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